ELSEVIER

Contents lists available at ScienceDirect

# **Bioorganic & Medicinal Chemistry Letters**

journal homepage: www.elsevier.com/locate/bmcl



# A new multi-gram synthetic route to labeling precursors for the $D_{2/3}$ PET agent $^{18}$ F-fallypride

Kwangho Kim, Nicole R. Miller, Gary A. Sulikowski, Craig W. Lindsley\*

Department of Chemistry and Pharmacology, Institute of Chemical Biology, Vanderbilt University, 802 Robinson Research Building, Nashville, TN 37235-1822, USA

## ARTICLE INFO

Article history: Received 21 June 2008 Revised 15 July 2008 Accepted 16 July 2008 Available online 20 July 2008

Keywords: PET D2 antagonist Imaging agent Fallypride

#### ABSTRACT

This Letter describes a new multi-gram synthetic protocol for the preparation of the classic tosylate labeling precursor for the  $D_{2/3}$  PET agent [ $^{18}$ F]fallypride. In the course of our studies, we also discovered two novel labeling precusors, the previously undescribed mesylate and chloro congeners of fallypride.

© 2008 Elsevier Ltd. All rights reserved.

Dysfunction in dopaminergic neurotransmission has been implicated in a number of neuropsychiatric disorders including Parkinson's disease, Alzheimer's disease, schizophrenia, and Huntington's disease. <sup>1-4</sup> The discovery and development of small molecule ligands that can selectively target dopamine receptors (D<sub>1</sub>–D<sub>4</sub>), transport, and synthesis is central to the development of therapeutic agents for the treatment of these complex diseases. <sup>1-4</sup> The D<sub>2</sub> receptor antagonists fallypride (FP) **1** and desmethoxy fallypride (DFP) **2** stand out due to their selectivity, affinity, and reversibility (Fig. 1). <sup>5,6</sup>

The corresponding [ $^{18}$ F] and [ $^{11}$ C]-labeled analogs of **1** have found utility as positron emission tomography (PET) agents. $^{7-13}$  In particular, [ $^{18}$ F]FP has been employed to study D $_{2/3}$  receptor occupancy and density in neuropsychiatric disorders and aging in both preclinical species and in humans. $^{7-15}$ 

The Vanderbilt Institute of Chemical Biology (VICB) established a Synthesis Core to provide synthetic and medicinal chemistry resources to the biomedical research community across the Vanderbilt campus, including the Vanderbilt Imaging Center. On one occasion, a request was made for a multi-gram synthesis of a labeling precursor for [18F]fallypride **4**, classically accessed by <sup>18</sup>F displacement of the corresponding tosylate **3** (Scheme 1).<sup>7–15</sup> Upon examination of the literature, we were dismayed to see that the classical published synthetic routes to **3** were performed on milligram quantities of material, employed preparative TLC for purification and afforded 30–50% yields for every step.<sup>8,9</sup> In 2007, Rosch and co-workers reported a large-scale synthesis utilizing a mixed

anhydride coupling between the corresponding substituted benzoic acid **5** and proline-derived amine **6** to afford **7** in an unspecified yield (Scheme 2).<sup>16</sup> Earlier de Paulis reported hydrob-

**Figure 1.** Structures of the  $D_{2/3}$  antagonists fallypride (FP), **1**, and desmethoxy fallypride (DFP), **2**.

**Scheme 1.** Synthesis of [<sup>18</sup>F]fallypride **4**. Reagents: (a) <sup>18</sup>F, Kryptofix, K<sub>2</sub>CO<sub>3</sub>, CH<sub>3</sub>CN.

<sup>\*</sup> Corresponding author. Tel.: +1 615 322 8700; fax: +1 615 343 6532. E-mail address: craig.lindsley@vanderbilt.edu (C.W. Lindsley).

**Scheme 2.** Existing routes to alcohol **7.** Reagents: (a) CICO<sub>2</sub>Et, Et<sub>3</sub>N; 9-BBN, H<sub>2</sub>O<sub>2</sub>, NaOH; (c) TFA, DCM; (d) allyl bromide, K<sub>2</sub>CO<sub>3</sub>, DMF.

oration–oxidation of allyl benzamide **8** to give alcohol **9** and following deprotection and N-allylation provide alcohol **7**.<sup>17</sup> In examining these synthetic routes, we became concerned with functional group compatibility under these reaction conditions and therefore turned our attention to developing a new synthetic route employing a protected derivative of benzoic acid **5** as a coupling partner.

To this end, alcohol **5** was derived from commercially available methyl benzoate 10 in three steps starting with O-methylation to afford benzoate 11 in 95% yield (Scheme 3). Hydroboration of 11 followed by treatment with basic peroxide then gave alcohol 12. Following ester hydrolysis (92%), benzoic acid 5 was treated with TBSCl and imidazole in DMF to provide TBS protected alcohol 13 in good overall yield. Next, the coupling of 13 with amine 6 (derived in two-steps from L-prolineamide) was examined. Optimal conditions employed EDCI in combination with HOBt and i-Pr2NEt as base to provide 14. Under these conditions, the coupled product was obtained in yields ranging from 45% to 64%. Finally, removal of the TBS group with TBAF delivered alcohol 7. Standard conditions then provided the tosylate 15, the immediate precursor to [18F]fallypride 4 in seven steps with an overall yield of 11%.<sup>18</sup> Importantly, all the steps leading to 14 were conducted on multigram scales with standard column chromatographic purification. while the final deprotection and tosylation steps were run on 1-2 gram scales (See Supporting Information).

The silyl protection of **5** and subsequent deprotection steps were a necessity for the multi-gram synthesis of **15**. All attempts to perform the amide coupling of **5** and **6**, with multiple peptide coupling reagents (EDCI, HATU, TFFH, etc) afforded poor results (yields less than 20%) and isolation of the extremely polar **7** proved difficult. Standard normal phase chromatography was insufficient on large scale to deliver pure **7** for the tosylation step. Thus, we were required to pursue preparative reverse phase chromatography with multiple injections, resulting in large volumes of  $CH_3CN/H_2O/TFA$  solutions to dry down with longer turn-around. While the protection/deprotection sequence adds two chemical steps, the overall yields and efficiency to access pure **15** are markedly improved.

The major problematic step in the synthesis was the amide coupling between the electron-rich, hindered benzoic acid **13** and

**Scheme 3.** Improved synthetic route to **15.** Reagents: (a) Mel, K<sub>2</sub>CO<sub>3</sub>, Me<sub>2</sub>CO, 95%; (b) i–9-BBN, THF, ii—NaOH, H<sub>2</sub>O<sub>2</sub>, 92%; (c) NaOH, THF (aq.), 92%; (d) TBSCl, ImH, DMF, 75%; (e) **6.** EDCI, HOBt, *i*-Pr<sub>2</sub>NEt, 45–64%; (f) TBAF, THF, 71%; (g) TsCl, DCM, pyridine, 44%.

Scheme 4. Synthesis of a novel labeling precursor CIFP 16 and fallypride 1. Reagents: (a) i-SOCl2, toluene, cat. DMF, ii-6, 31%; (b) TBAF, THF, reflux, 55%.

Scheme 5. Synthesis of a novel labeling precursor MsFP 17 and fallypride 1. Reagents: (a) i-MsCl, DCM, Et<sub>3</sub>N; (b) TBAF, t-BuOH, reflux, 40% over two steps.

amine **6**—a historically problematic reaction. We evaluated a number of coupling reagents (PyBop, BOP, TFFH), but none afforded advantages over EDCI. One attempted coupling that led to a new entity involved the treatment of benzoic acid **13** with thionyl chloride (Scheme 4). Reaction of the intermediate acid chloride with amine **6** led to the isolation of previously unknown chloride **16**, the result of substitution of the TBS ether for a chloro group, a variation of the Silyl-Durst chlorination. <sup>18,19</sup> This new entity **16** (CIFP) was evaluated as a D2 antagonist, and afforded an IC<sub>50</sub> of 17.1 nM. With **16** in hand, we then evaluated its ability to serve as a labeling precursor in route to [ $^{18}$ F]fallypride **4**. Exposure of **16** to TBAF in THF provided a 55% yield of **1**, indicating that this new CIFP congener **16** is a viable labeling precursor for [ $^{18}$ F]fallypride **4**. Activity at the D<sub>2</sub> receptor was also confirmed for our synthetic **1** with an IC<sub>50</sub> of 5.0 nM, a value in agreement with literature reports.

Based on these results, we examined the literature further, and were surprised to find that the corresponding mesylate analog **17** (MsFP) had never been prepared. To evaluate this potential labeling precursor, we intercepted alcohol **7** from the deprotection of **14** and treated it with MsCl in DCM and generated **17** (MsFP); however, this material was labile and proved difficult to isolate. Therefore, we generated **17** (MsFP) in situ and immediately treated the reaction mixture with TBAF in *t*-BuOH at 85 °C to afford a 40% yield of **1** (Scheme 5), suggesting that **17** (MsFP) could also serve as a viable labeling precursor for the preparation of [<sup>18</sup>F]fallypride **4**.<sup>18</sup>

In summary, we have developed an improved, high yielding, and scalable synthetic route to the classical labeling precursor **15** for [ $^{18}$ F]fallypride **4**. During the course of this work, we discovered **16** (CIFP), a novel  $D_2$  antagonist and viable labeling precursor for [ $^{18}$ F]fallypride **4**. This work also prepared and evaluated the previously unknown mesylate congener, **17** (MsFP), as a labeling precursor, and found that it too was viable. Further studies in this arena are in progress and will be reported in due course.

# Acknowledgments

The authors thank the Vanderbilt Institute of Chemical Biology, the Vanderbilt Imaging Center (Dr. Ron Baldwin), and the Vanderbilt Department of Pharmacology for support of this research.

#### Supplementary data

Experimental procedures and analytical data for compounds **1**, **11-17** are provided. This material is available free of charge via the Internet at http://www.sciencedirect.com/science/journal/0960894X. Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.bmcl.2008.07.065.

## References and notes

- 1. Stoof, J. C.; Kebabian, J. W. Life Sci. 1984, 35, 2281.
- Carbon, M.; Ghilardi, M. F.; Fukuda, M.; Silvestri, G.; Mentis, M. J.; Ghez, C.; Moeller, J. R.; Eidelberg, D. Hum. Brain Mapp. 2003, 19, 197.
- 3. Chesselet, M. F.; Delf, J. M. *Trends Neurosci.* **1996**, 19, 417.
- 4. Seeman, P.; Bzowej, N. H.; Guan, H. C.; Bergeron, C.; Reynolds, G. P.; Bird, E. D.; Riederer, P.; Jellinger, K.; Tourtellotte, W. W. Neuropsychopharmacology 1987, 1, 5.
- 5. Grunder, G.; Siessmeier, T.; Piel, M.; Vernaleken, I.; Buchholz, H. G.; Zhou, Y.; Hiemke, C.; Wong, D. F.; Rosch, F.; Bartenstein, P. J. Nuc. Med. 2003, 44, 109.
- Siesssmeier, T.; Zhou, Y.; Buchholz, H-G.; Landvogt, C.; Vernaleken, I.; Piel, M.; Schirrmacher, R.; Rosch, F.; Schreckenberger, M.; Wong, D. F.; Cummings, P.; Grunder, G.; Barnstein, P. J. Nuc. Med. 2005, 46, 964.
- Grunder, G.; Landvogt, C.; Vernaleken, I.; Buchholz, H. G.; Ondraeck, J.; Siesssmeier, T.; Hartter, S.; Schreckenberger, M.; Stoeter, P.; Hiemke, C.; Rosch, F.; Wong, D. F.; Bartenstein, P. Neuropsychopharmacology 2006, 31, 1027.
- 8. Mukherjee, J.; Shi, B. Z.; Christian, B. T.; Chattopadhyay, S.; Narayanan, T. K. Bioorg. Med. Chem. 2004, 12, 95.
- Mukherjee, J.; Yang, Z-Y.; Das, M. K.; Brown, T. Nuc. Med. Biol. 1995, 22, 283.
- 10. Mukherjee, J. J. Fluorine Chem. 1990, 49, 151.
- (a) Mukherjee, J. Int. J. Appl. Radiat. Isotopes 1991, 42, 713; (b) Mukherjee, J.;
  Yang, Z-Y.; Copper, M. Eur. J. Pharmacol. 1992, 175, 363.
- Mathis, C. A.; Bishop, J. E.; Gerdes, J. M.; Whitney, J. M.; Brennan, K. M.; Jagust, W. J. Nuc. Med. Biol. 1992, 19, 571.
- (a) Kessler, R. M.; Mason, N. S.; de Paulis, T.; Ansari, M. S.; Schmidt, D.; Manning, R. G.; Votaw, J. R. *J. Nuc. Med.* **1992**, 33, 847; (b) Kessler, R. M.; de Paulis, T.; Ansari, M. S.; Schmidt, D.; Manning, R. G.; Votaw, J. R. *J. Nuc. Med.* **1993**, 34, 202P.
- Mukherjee, J.; Christian, B. T.; Narayanan, T. K.; Shi, B.; Collins, D. Brain Res. 2005, 1032, 77.
- Riccardi, P. R.; Li, M. S.; Ansari, D.; Zald, S.; Park, B.; Dawant, S.; Anderson, M.; Doop, N.; Woddward, E.; Schoenberg, D.; Schmidt, R.; Baldwin, R.; Kessler, R. Neuropsychopharmacology 2006, 31, 1016.
- Stark, D.; Piel, M.; Hubner, H.; Gmeiner, P.; Grunder, G.; Rosch, F. Bioorg. Med. Chem. 2007, 15, 6819.
- 17. de Paulis, T. Curr. Pharmaceut. Design 2003, 9, 673.
- 18. For full experimental details, see the Supplementary Data.
- 19. Oddon, G.; Ugen, U. Tetrahedron Lett. 1997, 25, 4407.